

# NEW SIPHULIN DERIVATIVES FROM THE LICHEN *SIPHULA CERATITES*

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**Key Word Index**—*Siphula ceratites*; Usneaceae; siphulin; protosiphulin; oxysiphulin; lichen metabolites; chromenones.

**Abstract**—From the lichen, *Siphula ceratites*, two new metabolites, protosiphulin and oxysiphulin, were isolated besides the known compound, siphulin. The structures of the two new chromenones were proved by  $^1\text{H}$  NMR and  $^{13}\text{C}$  NMR spectral analysis in comparison with those of siphulin.

The lichen of *Siphula* species are mostly distributed on the Southern hemisphere, and only *Siphula ceratites* (Wahlenb.) Fr. has been found on the northern part of the Scandinavian peninsula and Canada. This lichen is noted chemotaxonomically since it contains a characteristic chromomenone-type compound, siphulin (1), which has never been found in any other species of *Siphula* collected on the Southern hemisphere [1, 2].

From acetone extracts of *Siphula ceratites* collected in Canada, 2 new compounds 2 and 3 have been

isolated along with siphulin by chromatographic separation. 2,  $\text{C}_{24}\text{H}_{28}\text{O}_8$ , was readily converted into siphulin, on treating with methanolic HCl. Therefore, 2 must be a hydrated derivative of siphulin 1. This assumption was supported by the following NMR spectral comparison of 2 with 1.

The signal  $\delta$  5.74 (1H) assigned to a proton at  $\text{C}_{(3)}$  of 1 was not observed in the  $^1\text{H}$  NMR of 2, whereas signals giving a geminal coupling at  $\delta$  3.00 and 3.32 (1H each,  $d$ ,  $J = 16$  Hz;  $\text{CH}_2$ ) assigned to protons at  $\text{C}_{(3)}$  as well as a higher shifted singlet at  $\delta$  3.45 (2H) assigned to  $\text{C}_{(7)}$  were given by 2.

The  $^{13}\text{C}$  NMR spectra of 1 and 2 were also compared (Table 1) with those of flavones and flavanones [3]. The  $^{13}\text{C}$  NMR signal assigned to  $\text{C}_{(4)}$  of flavanone shows a downfield shift in comparison with that of flavone. The same tendency was observed in 2 in

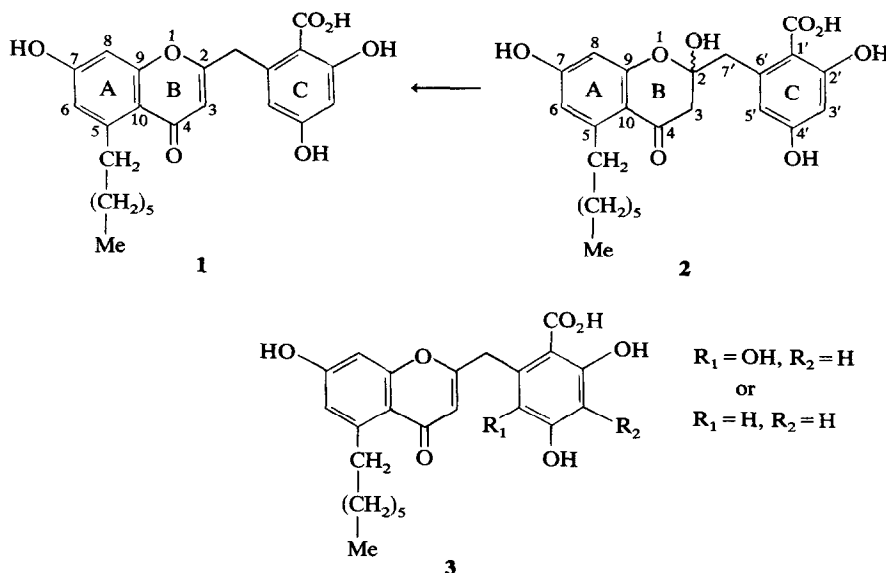


Table 1.  $^{13}\text{C}$  chemical shifts ( $\delta$  ppm) of acacetin and isosakuranetin in comparison with those of siphulin (**1**) and protosiphulin (**2**)

Compound	Carbon No.								
	C <sub>(2)</sub>	C <sub>(3)</sub>	C <sub>(4)</sub>	C <sub>(5)</sub>	C <sub>(6)</sub>	C <sub>(7)</sub>	C <sub>(8)</sub>	C <sub>(9)</sub>	C <sub>(10)</sub>
Acacetin	168.9	103.9	182.3	157.9	99.4	164.8	94.3	162.2	104.4
(5,7-Dihydroxy 4'-methoxy flavone)									
<b>1</b>		116.9	179.9						
Isosakuranetin	77.8	41.9	196.1	163.4	95.7	166.6	94.8	162.8	101.7
(5,7-Dihydroxy 4'-methoxy flavanone)									
<b>2</b>	100.4	47.9	188.4						

comparison with **1** to show that a double bond is absent from the B ring.

A signal at  $\delta$  47.9 in the  $^{13}\text{C}$  NMR of **2** which gave a triplet on off-resonance was assigned to C<sub>(3)</sub>, since one of the five doublet signals around 110 ppm on off-resonance of the  $^{13}\text{C}$  NMR spectrum of **1** disappeared in that of **2**. The signal at  $\delta$  100.4 given by **2** was assigned to the quaternary C<sub>(2)</sub>, since it is the only singlet signal which appeared at a higher field and was not observed in the spectrum of **1**. The singlet signals at  $\delta$  104.8 and 115.0 of  $^{13}\text{C}$  NMR of **1** and the corresponding signals at  $\delta$  105.8 and 112.7 of **2** were assigned to the aromatic carbons attached to carbonyls. A quartet  $^{13}\text{C}$  signal which appeared at  $\delta$  14.2 for both **1** and **2** was assigned to a terminal methyl of the alkyl side chain, and a triplet at  $\delta$  35.7 for both **1** and **2** was assigned to benzyl methylene of the side chain. Other triplets in the region of  $\delta$  23.2~37.3 for **1** and  $\delta$  23.3~37.3 for **2** were represented by other methylenes of the alkyl side chain. The doublet signals in the region of  $\delta$  101.7~115.0 for **1** and  $\delta$  102.3~114.1 for **2** accounted for the unsubstituted aromatic ring carbons, and the singlets appeared in the region of  $\delta$  140.7~173.2 for **1** and  $\delta$  139.8~167.8 for **2** were assigned to other substituted aromatic carbons. Formulation of the structure of **2** is thus complete.

Compound **3**, C<sub>24</sub>H<sub>26</sub>O<sub>8</sub>, gave almost the same  $^1\text{H}$  NMR spectrum as **1**, except that there is a singlet at  $\delta$  6.72 (1H) instead of at 6.66 (2H) with **1**. This revealed the presence of one more hydroxyl substituted in ring C of **3**. The hydroxyl substitution in **3** was also supported by its greyish-violet colouration with FeCl<sub>3</sub> in contrast with the reddish-purple colour given by **1** and **2**. The *R<sub>f</sub>* value of **3** (0.2) on TLC was lower than that of **1** (0.3). The high resolution MS of **3** gave a highest mass peak at 398.1728 (calc. for C<sub>23</sub>H<sub>26</sub>O<sub>6</sub>, M-CO<sub>2</sub>: 398.1786) and base peak at 327.0867 (calc. for C<sub>18</sub>H<sub>15</sub>O<sub>6</sub>: 327.0852), while **1** gave 382.1754 (calc. for C<sub>23</sub>H<sub>26</sub>O<sub>5</sub>, M-CO<sub>2</sub>: 382.1778) and 311.0976 (calc. for C<sub>18</sub>H<sub>15</sub>O<sub>5</sub>: 311.0918).

Consequently **3** must have the structure shown, except that conclusive evidence for the position of the hydroxyl, R<sub>1</sub> or R<sub>2</sub>, has not been obtained due to the shortage of material.

From the biogenetic viewpoint **2** would be an immediate precursor of siphulin **1**, so that it is designated protosiphulin, while **3** is named oxysiphulin as it is an oxygenated derivative of siphulin.

#### EXPERIMENTAL

IR spectra were measured in KBr;  $^1\text{H}$  NMR and  $^{13}\text{C}$  NMR spectra were recorded in Me<sub>2</sub>CO-*d*<sub>6</sub> with TMS as int. standard.

**Extraction and isolation.** Lichen material, *Siphula ceratites* (Wahlenb.) Fr. (305 g) collected in British Columbia, Canada, was extracted with Me<sub>2</sub>CO. On concn siphulin was precipitated to separate supernatant which was chromatographed over a 0.5 N oxalic acid-impregnated Si gel column eluting with C<sub>6</sub>H<sub>6</sub>-Me<sub>2</sub>CO and then over a Sephadex LH-20 eluted with Me<sub>2</sub>CO, siphulin (**1**) (total yield: 6.38 g, 2.1%), protosiphulin (**2**) (yield: 537 mg, 0.17%) and oxysiphulin (**3**) (yield: 18 mg, 0.006%).

**Siphulin (1).** Colourless needles, mp 179–181° (decomp.) (from aq. MeOH); UV and IR as in lit. [2];  $^1\text{H}$  NMR (in Me<sub>2</sub>CO-*d*<sub>6</sub>) ppm:  $\delta$  0.86 (3H, *m*, CH<sub>3</sub>-(CH<sub>2</sub>)<sub>5</sub>-CH<sub>2</sub>-), *ca.* 1.3 (10H, *m*, Me(CH<sub>2</sub>)<sub>5</sub>-CH<sub>2</sub>-), 3.16 (2H, *tr*, *J* = 8 Hz, Me-(CH<sub>2</sub>)<sub>5</sub>-CH<sub>2</sub>-), 4.26 (2H, *s*, -CH<sub>2</sub>- at 7'), 5.74 (1H, *s*, C<sub>(3)</sub>-H), 6.33 (1H, *d*, *J* = 2 Hz, C<sub>(6 or 8)</sub>-H), 6.42 (1H, *d*, *J* = 2 Hz, C<sub>(6 or 8)</sub>-H), 6.66 (2H, *s*, C<sub>(3')</sub>-H and C<sub>(5')</sub>-H);  $^{13}\text{C}$  NMR (in Me<sub>2</sub>CO-*d*<sub>6</sub>) ppm:  $\delta$  14.2 (CH<sub>3</sub>-(CH<sub>2</sub>)<sub>5</sub>-CH<sub>2</sub>), 35.7 (Me-(CH<sub>2</sub>)<sub>5</sub>-CH<sub>2</sub>-), 40.4 (C<sub>(7')</sub>), 116.9 (C<sub>(3)</sub>), 148.1 (C<sub>(6')</sub>), 179.9 (C<sub>(4)</sub>). MS: 382.1754 (M-CO<sub>2</sub>), 339.1246 (382-Me-CH<sub>2</sub>-CH<sub>2</sub>), 325.1069 (339-CH<sub>2</sub>), 311.0976 (325-CH<sub>2</sub>) (base peak); Calc. for C<sub>23</sub>H<sub>26</sub>O<sub>5</sub> (M-CO<sub>2</sub>): 382.1778, C<sub>20</sub>H<sub>16</sub>O<sub>5</sub>: 339.1231, C<sub>10</sub>H<sub>17</sub>O<sub>5</sub>: 325.1074, C<sub>18</sub>H<sub>15</sub>O<sub>5</sub>: 311.0918.

**Protosiphulin (2).** Colourless needles, mp 179–182° (decomp.) (from Me<sub>2</sub>CO-hexane); [ $\alpha$ ]<sub>D</sub><sup>20</sup> ± 0°; UV  $\lambda_{\text{max}}^{\text{EtOH}}$  nm (log  $\epsilon$ ): 230 (4.41), 278 (4.41); IR  $\nu_{\text{max}}^{\text{KBr}}$  cm<sup>-1</sup>: 3300, 1663, 1655, 1650, 1635, 1603;  $^1\text{H}$  NMR (in Me<sub>2</sub>CO-*d*<sub>6</sub>) ppm:  $\delta$  0.89 (3H, CH<sub>3</sub>-(CH<sub>2</sub>)<sub>5</sub>-CH<sub>2</sub>-), *ca.* 1.3 (10H, *m*, Me-(CH<sub>2</sub>)<sub>5</sub>-CH<sub>2</sub>-), 3.00 (1H, *d*, *J* = 16 Hz, C<sub>(3)</sub>-H), 3.1 (2H, *tr*, *J* = *ca.* 8 Hz, Me-(CH<sub>2</sub>)<sub>5</sub>-CH<sub>2</sub>-), 3.32 (1H, *d*, *J* = 16 Hz, C<sub>(3)</sub>-H), 3.45 (2H, *s*, CH<sub>2</sub> at 7'), 6.11 (1H, *d*, *J* = 2 Hz, C<sub>(6 or 8)</sub>-H), 6.28 (1H, *d*, *J* = 2 Hz, C<sub>(6 or 8)</sub>-H), 6.40 (1H, *d*, *J* = 2 Hz, C<sub>(3' or 5')</sub>-H), 6.42 (1H, *d*, *J* = 2 Hz, C<sub>(3' or 5')</sub>-H), 9.4 (1H, *s*, OH) 10.80 (1H, *s*, OH);  $^{13}\text{C}$  NMR (in Me<sub>2</sub>CO-*d*<sub>6</sub>) ppm:  $\delta$  14.2 (CH<sub>3</sub>-(CH<sub>2</sub>)<sub>5</sub>-CH<sub>2</sub>-), 35.6 (Me-(CH<sub>2</sub>)<sub>5</sub>-CH<sub>2</sub>-), 37.2 (C<sub>(7')</sub>), 47.9 (C<sub>(3)</sub>), 100.4 (C<sub>(2)</sub>), 149.5 (C<sub>(6')</sub>), 188.4 (C<sub>(4)</sub>); MS: 382.1829 (M-H<sub>2</sub>O-CO<sub>2</sub>), 339.1266 (382-Me-CH<sub>2</sub>-CH<sub>2</sub>), 325.1163 (339-CH<sub>2</sub>),

311.0839 (325-CH<sub>2</sub>) (base peak). Calc. for C<sub>23</sub>H<sub>26</sub>O<sub>5</sub> (M-H<sub>2</sub>O-CO<sub>2</sub>): 382.1778, C<sub>20</sub>H<sub>19</sub>O<sub>5</sub>: 339.1231, C<sub>19</sub>H<sub>17</sub>O<sub>5</sub>: 325.1075, C<sub>18</sub>H<sub>15</sub>O<sub>5</sub>: 311.0917. The soln of **2** in MeOH containing a drop of conc HCl was refluxed for 10 min at 100°. The product was purified through Sephadex LH 20 using Me<sub>2</sub>CO as the solvent. The product was siphulin (**1**) identified by TLC, IR, NMR and MS comparison.

**Oxysiphulin (3)**. Colourless needles, mp 176-178° (decomp.) (from aq. MeCO); UV λ<sub>max</sub><sup>EtOH</sup> nm (log ε): 233 (sh) (4.46), 290 (4.18); <sup>1</sup>H NMR (in Me<sub>2</sub>CO-d<sub>6</sub>) ppm: δ 0.83 (3H, CH<sub>3</sub>-(CH<sub>2</sub>)<sub>5</sub>-CH<sub>2</sub>), ca. 1.3 (10H, m, Me-(CH<sub>2</sub>)<sub>5</sub>-CH<sub>2</sub>), 3.31 (2H, tr, Me-(CH<sub>2</sub>)<sub>5</sub>-CH<sub>2</sub>-), 4.42 (2H, s, CH<sub>2</sub> at 7'), 5.67 (1H, s, C<sub>(3)</sub>-H), 6.30 (1H, d, J = 2 Hz, C<sub>(6 or 8)</sub>-H), 6.38 (1H, d, J = 2 Hz, C<sub>(6 or 8)</sub>-H), 6.72 (1H, s, C<sub>(3' or 5')</sub>-H). MS: 398.1728 (M-CO<sub>2</sub>), 355.1180 (398-Me-CH<sub>2</sub>-CH<sub>2</sub>-), 341.1033 (355-CH<sub>3</sub>), 327.0867 (341-CH<sub>2</sub>) (base peak), 313.0711 (327-CH<sub>2</sub>). Calc. for C<sub>23</sub>H<sub>26</sub>O<sub>6</sub>(M-CO<sub>2</sub>): 398.1786, C<sub>20</sub>H<sub>19</sub>O<sub>6</sub>: 355.1103, C<sub>19</sub>H<sub>17</sub>O<sub>6</sub>: 341.1034, C<sub>18</sub>H<sub>15</sub>O<sub>6</sub>: 327.0852, C<sub>17</sub>H<sub>13</sub>O<sub>6</sub>: 313.0764.

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## EIN NEUES SYRINGAALKOHOL-DERIVAT AUS *ERECHTITES HIERACIFOLIA*

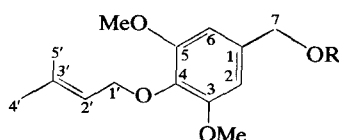
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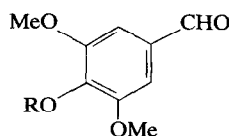
(Eingegangen am 8 Juni 1979)

**Key Word Index**—*Erechtites hieracifolia*; Compositae; new syringyl alcohol derivative.

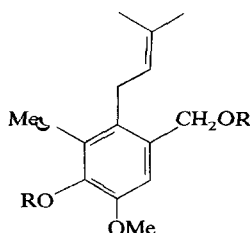
Aus *E. Hieracifolia* Raf. (Tribus Senecioneae) sind bisher nur Pyrrolizidin-Alkaloide isoliert worden [1], wie sie offenbar für grosse Teile der Tribus Senecioneae charakteristisch sind. Wir haben jetzt die Neutralstoffe näher untersucht. Die Wurzeln enthalten neben β-Farnesen und Bicyclogermacren einen Angelicaester, dem die Konstitution **1** zukommt wie aus den spektroskopischen Daten weitgehend sichergestellt werden kann (s. Tabelle 1). Obwohl biogenetisch die vicinale Anordnung der drei O-Funktionen wahrscheinlich war, haben wir diese durch Synthese des entsprechenden Acetats sichergestellt. Durch Alkylierung von Syringaaldehyd (**4**) mit 3,3-Dimethylallylbromid erhält man der Ether **5** und daraus mit Alanat den Alkohol **2**, der mit Acetanhydrid das Acetat **3** liefert. Die <sup>1</sup>H-NMR-Daten (s. Tabelle 1) von **3** sind praktisch identisch mit denen von **1**. Lediglich die Lage der CH<sub>3</sub>-Gruppe ist wie in ähnlichen Fällen geringfügig verschieden. Bemerkenswert ist eine sehr leicht verlaufende säurekatalysierte Umlagerung von **2**, die zu **6** führt, das ebenfalls als Diacetat (**1**) isoliert wurde. (<sup>1</sup>H-NMR-Daten siehe Tabelle 1). Bei der Alanat-Reduktion von **5** ist daher nach Zersetzen mit NH<sub>4</sub>Cl Neutralwaschen mit Natriumhydrogencarbonat unerlässlich.



- 1** R = Ang  
**2** R = H  
**3** R = Ac



- 4** R = H  
**5** R = CH<sub>2</sub>CH=CM<sub>2</sub>



- 6** R = H  
**7** R = Ac